© Mary Ann Liebert, Inc. DOI: 10.1089/aid.2021.0173

> Open camera or QR reader and scan code to access this article and other resources online.



A Summary of the Sixth International Workshop on Microbiome in HIV Pathogenesis, Prevention, and Treatment

Scott Sherrill-Mix,^{1,i} Michelle Yang,² Grace M. Aldrovandi,³ Jason M. Brenchley,⁴ Frederic D. Bushman,¹ Ronald G. Collman,⁵ Satya Dandekar,⁶ Nichole R. Klatt,⁷ Laurel A. Lagenaur,⁸ Alan L. Landay,⁹ Roger Paredes,¹⁰ Gilda Tachedjian,¹¹ Jim A. Turpin,¹² Sergio Serrano-Villar,¹³ Catherine A. Lozupone,¹⁴ and Mimi Ghosh²

Abstract

In October of 2020, researchers from around the world met online for the sixth annual International Workshop on Microbiome in HIV Pathogenesis, Prevention, and Treatment. New research was presented on the roles of the microbiome on immune response and HIV transmission and pathogenesis and the potential for alterations in the microbiome to decrease transmission and affect comorbidities. This article presents a summary of the findings reported.

Keywords: HIV/SIV, microbiome, pathogenesis, therapeutics, comorbidities, transmission, prevention

Introduction

THIS YEAR MARKED the sixth meeting of the International THIS YEAR MARKED the SIXUI Include of the Sixui Presence aims to bring vention, and Treatment. 1-5 The conference aims to bring together experts from across the field of microbiome research for a multidisciplinary discussion of the impact of the microbiome on HIV outcomes. Due to the COVID-19 pandemic, the conference was held virtually.

In spite of the difficulties, researchers came together on the virtual platform to discuss interesting findings on the role of the microbiome and inflammation in HIV-1 transmission, prevention, and comorbidities. The first keynote address by Dr. Elodie Ghedin described her work to identify changes in bacteriophage and host bacteria during viral infection, while the second keynote address by Dr. Cynthia Sears discussed her work identifying links between the gut microbiome and colorectal cancer.

First Keynote Address

The 2020 conference opened with a keynote presentation on the dynamics of phage-bacteria interactions in the respiratory tract from Dr. Elodie Ghedin from the National

Department of Microbiology, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA.

Department of Epidemiology, The George Washington University, Washington, District of Columbia, USA.

Department of Pediatrics, University of California, Los Angeles, California, USA.

⁴Laboratory of Viral Diseases, NIAID, NIH, Bethesda, Maryland, USA.

⁵Department of Medicine, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA.

Department of Medical Microbiology and Immunology, University of California, Davis, Davis, California, USA.

Department of Medicine, University of Minnesota, Minnesota, USA.

⁸Osel, Inc., Mountain View, California, USA.

Division of Gerontology, Department of Internal Medicine, Rush University Medical Center, Chicago, Illinois, USA.

¹⁰Institut de Recerca de la SIDA IrsiCaixa i Unitat VIH, Universitat Autònoma de Barcelona, Universitat de Vic, Catalonia, Spain.

¹¹ Burnet Institute, Melbourne, Australia.
12 Divison of AIDS, NIAID, NIH, Bethesda, Maryland, USA.

¹³Department of Infectious Diseases, Hospital Universitario Ramon y Cajal, Madrid, Spain.

¹⁴Department of Medicine, University of Colorado Denver, Aurora, Colorado, USA.

ORCID ID (https://orcid.org/0000-0002-8955-8072).

Institute of Allergy and Infectious Diseases. Phages have a multitude of effects on the microbiota and, in addition, can interact with immune cells directly. Dr. Ghedin's team investigated households in Nicaragua with index cases positive for influenza and asked whether there was higher transmission of pathogenic bacteria and antibiotic resistance genes in households with higher rates of intrahousehold influenza transmission. Metatranscriptome and metagenome analyses of nasal washes showed both differential abundance and differential expression of bacteria and phages that had some association with the influenza status of the subject.

The team also applied a novel method to investigate phage-bacteria interaction by matching the phages they observed to sequences detected in bacterial CRISPR arrays. The CRISPR system occurs in many bacterial species and serves as an acquired antiviral defense by incorporating DNA sequence from previous phage infections into a CRISPR array that is then used to guide cleavage of any future DNA matching that sequence. Combining abundance-based and CRISPR-based inference of phage-bacteria interactions enables high-resolution capture of dynamic interactions in the microbiome and is being used to further characterize transmission of bacteria within and between households.

Transmission

The progestin-based contraceptive depot medroxyprogesterone acetate (DMPA) has been reported to be associated with increased risk of HIV acquisition and transmission in multiple observational studies. 10 In contrast, the recent Evidence for Contraceptive Options and HIV Outcomes Trial¹¹ reported no increased risk of HIV acquisition in women using DMPA. However, the interpretation of these data remain controversial as the study did not have a "no contraceptive" control group and the study was designed to detect a 50% or greater increase in HIV acquisition risk. 12-15 Multiple studies have reported changes in genital immune parameters among DMPA users, ¹⁶ although, no major changes in microbiome have been described in the majority of these studies. 17,18 Increased risk of HIV acquisition has, however, been described in DMPA users who have a polymicrobial vaginal microbiome, a known risk factor for increased acquisition of HIV.19,20

To understand the mechanisms underlying this association, Dr. Adam Burgener from Case Western Reserve University used previously published microbiome data from the CAPRISA-004 trial²¹ and compared the relative risk of HIV acquisition among those using DMPA, norethisterone enanthate (NET-EN), and combined oral contraceptives (COC). They found that in women whose vaginal microbiome was Lactobacillus dominant, HIV acquisition was threefold higher in women using DMPA relative to women using NET-EN or COC, whereas women whose microbiomes were not Lactobacillus dominant showed no significant difference.²² Higher serum medroxyprogesterone acetate levels associated with increased molecular markers of inflammation in the vaginal mucosal fluid of Lactobacillus-dominant women while women who had non-Lactobacillus-dominant microbiomes displayed markers of inflammation regardless of contraceptive use.²² Although limited by the nonrandomized post hoc design of the study, these findings nonetheless suggest an interaction among microbiome, hormonal contraceptives, and HIV susceptibility.

Dr. Colleen Kelley from Emory University discussed the immune environment of the rectal mucosa and the impact on HIV transmission. In the United States, men who have sex with men (MSM) are the group most highly affected by HIV-1 with the majority of transmission occurring through condomless receptive anal intercourse (CRAI). 23,24 The rectal epithelium, distinct from vaginal and penile epithelium, is composed of a single layer of cells that results in a fragile barrier that offers limited protection to viral entry. Furthermore, the gastrointestinal tract is rich in CD4+CCR5+ cells, the primary targets of HIV. Dr. Kelley and collaborators have previously reported distinct rectal mucosal transcriptomes, including enrichment in immune activation and mucosal injury and repair pathways, and changes in the microbiome, including an enrichment of Prevotella, in MSM who practice CRAI.²⁵ Their ongoing studies have shown an association of CRAI with markers of cellular proliferation and neutrophil infiltration of crypt epithelial cells in rectal mucosa, but not with gut microbiome. ²⁶ In MSM taking oral pre-exposure prophylaxis, no shift in microbiota was observed.²⁷ However, the use of hyperosmolar lubricants (not uncommon for CRAI) resulted in a shift in microbiome from Bacteroides to Prevotella.27 Ongoing studies also show that adolescent MSM have distinct rectal immune microenvironment and microbiome compared with adult MSM with samples from young MSM showing higher Bacteroides to Prevotella ratios and higher HIV replication in an explant challenge model.

Dr. Jason Brenchley from the National Institute of Allergy and Infectious Diseases discussed his findings about the association of gut microbiome and simian immunodeficiency virus (SIV) acquisition in Asian macaques. There is little evidence for dysbiosis induced by SIV infection in macaques.²⁸ Similarly, many changes in human gut microbiota initially linked to HIV infection appear to instead associate with sexual practices.^{29,30} The effects of these externally caused microbial dysbioses on HIV pathogenesis and transmission remain uncertain. The Brenchley laboratory disrupted the microbiome and induced intestinal epithelial damage in macaques using vancomycin then infected them with SIV and found their progression to AIDS was similar to untreated macaques.³¹ Vancomycin treatment did appear to induce low-level gastrointestinal tract inflammation and result in an increase in the number of transmitted founder viruses establishing new infections during rectal SIV challenge. To move beyond changes in the bacteria of the gut, the Brenchley laboratory is also working to viably culture translocating bacteria from liver, lymph nodes, and spleen and has identified a methyl transferase potentially shared among many translocating species.

Pathogenesis

MSM have an altered gut microbiome, elevated immune activation, and CCR5 expression even in the absence of HIV infection. ^{25,29,30,32,33} To investigate this phenomenon, Dr. Eiko Yamada from the University of Colorado presented data showing that fecal microbes, especially *Holdemanella biformis*, a strain enriched in the guts of MSM, can upregulate

CCR5 expression and alter tumor necrosis factor- α /interleukin (IL)-10 ratio, concordant with previous observations of a proinflammatory role for *H. biformis*.³⁴

Dr. Yanhui Cai from Gilead Sciences tested fecal microbiome from virologically suppressed HIV-infected individuals and observed no significant differences compared with healthy volunteers. However, the abundance of Proteobacteria in infected individuals correlated with interferonstimulated gene expression and CD4 T cell activation.

Dr. Shilpa Ray from the Karolinska Institute presented microbiome data on a longitudinal cohort of HIV-infected individuals before and after initiation of several antiretroviral therapy (ART) regimens. They observed relatively little change in bacterial diversity or richness following ART initiation when all ART regimens were pooled but significant decreases in alpha diversity when only patients treated with non-nucleoside reverse transcriptase inhibitor-based therapies were considered. To follow up on this finding, they tested the antibacterial properties of various antiretroviral drugs and found that zidovudine and efavirenz inhibited the growth of several bacterial species, showing the potential for ART to directly affect the microbiome.³⁵

James Virga from the National Cancer Institute reported results from the GUTCHEK study, where the effects of oral antibiotic rifaximin³⁶ on gut inflammation, immune activation, and microbiome were investigated in people living with HIV on long-term ART. Although a significant decrease in total diversity was observed in fecal samples following rifaximin treatment, no significant changes in soluble inflammatory markers, immune cell subsets, or plasma HIV RNA were observed after rifaximin treatment.

Second Keynote Address

In the second keynote presentation, Dr. Cynthia Sears from John Hopkins University discussed her work studying the links between the gut microbiome and colorectal cancer. Rates of colorectal cancer are rising around the world, especially in relatively younger age groups. Metagenomic sequencing has revealed ties between cancer and the microbial communities of the gut offering the opportunity for characterization of the microbiome to inform prevention and develop therapies. For example, enterotoxigenic *Bacteroides fragilis* express a toxin that can induce DNA damage, trigger inflammation pathways, and lead to carcinogenesis. Another sign of a microbial link is that biofilms are often found in association with colorectal tumors.

To test the causality of this association, the Sears laboratory and collaborators transferred biofilm-containing microbiota from homogenized biopsies from colorectal cancer patients by gavage to germ-free mouse models of carcinogenesis and observed subsequent biofilm formation in the distal colon, immune cell infiltration into lamina propia, and tumorigenesis. In an unpublished work, the laboratory has cultured bacterial isolates from a patient sample and recapitulated mouse tumorigenesis using a combination of 30 isolates. Further work in patients with familial adenomatous polyposis showed a higher-than-expected frequency of colonic biofilms containing bacteria expressing oncogenic toxins. So

The Sears Laboratory has also analyzed databases of clinical and sequencing data to probe links between host and microbiome. Using electronic health records from the United

Kingdom, they showed associations between antibiotic use and colorectal cancer diagnosis in a matched case—control study. ⁵¹ By careful reprocessing of raw bacterial sequencing data from multiple studies, meta-analyses identified an enrichment of several bacterial taxa in the microbiomes of colorectal cancer patients ⁴⁸ and decreased diversity associated with HIV infection in woman and men who have sex with women but not in MSM ⁵² with ongoing analysis to distinguish the effects of HIV infection from those of sexual orientation.

Lessons learned from these analyses included the need for studies to provide sufficient metadata, the potential confounding factor of technical differences between laboratories⁵³ and the opportunity to integrate data on chemical, social, and environmental exposures.⁵⁴

Prevention

A *Lactobacillus*-dominated vaginal microbiome appears to reduce the risk of acquiring HIV while more diverse communities and bacterial vaginosis associate with an increased risk. ^{55–57} However, antibiotic treatment of bacterial vaginosis has low cure rates and results in frequent recurrence. ^{58–60}

Dr. Laurel Lagenaur from Osel, Inc., reported on the results of a phase 2b clinical trial (NCT02766023) of the treatment of bacterial vaginosis aided by the a biotherapeutic formulation of a naturally occurring *Lactobacillus crispatus* strain CTV-05^{61,62} called LACTIN-V. The study showed *L. crispatus* colonization in most women treated with LACTIN-V and a marked decrease in bacterial vaginosis recurrence over antibiotics alone. ^{63,64} Colonization was not affected by condom use or menses and no severe adverse events were observed. A subsequent phase 3 trial is currently being discussed with the FDA.

To better understand the link between lactic acid-producing *Lactobacillus* in the vaginal microbiome and decreased HIV acquisition, ^{65,66} Brianna Jesaveluk from the Burnet Institute discussed her research testing the effects of lactic acid in a cell model of barrier integrity. Measurements of transepithelial electrical resistance revealed that lactic acid at an acidic pH increased barrier integrity, whereas controls of hydrochloric acid at a similarly acidic pH or lactic acid at a neutral pH did not. To investigate this difference, RNA-Seq was used to characterize gene expression in the cell line with and without lactic acid and confirmed that several tight junction genes appeared upregulated in the presence of lactic acid.

Kaitlin Marquis from the University of Pennsylvania reported on her attempts to discover additional metabolites linking the vaginal microbiome to HIV acquisition risk. Using a meta-analysis of previously published studies of metabolites, ^{67–70} she found a set of metabolites enriched in high-diversity vaginal communities that have been linked to HIV acquisition. To follow up on these candidates, she developed a high-throughput screen using a reporter cell line to assess the effects of over 500 metabolites on *in vitro* HIV replication and found several candidate metabolites, including 2-hydroxyisovalerate that were both enriched in high-diversity communities and promoted HIV replication.

Male circumcision has been linked to a reduced risk of acquiring HIV.^{71–74} Dr. Rupert Kaul from the University of Toronto described his work to study how the foreskin increases HIV risk and whether there are nonsurgical

alternatives to reduce this risk. Circumcision reduces inflammatory cytokines⁷⁵ and changes the penile microbiome⁷⁶ with a reduction in the abundance of several anaerobic bacterial taxa associated with inflammation and HIV risk.⁷⁷ Dr. Kaul reported on ongoing work showing that unlike the vaginal microbiome there do not appear to be clear penile community types or protective bacterial taxa but that several bacterial species were associated with HIV seroconversion. The associations of penile microbiome and HIV acquisition suggest the possibility that antibiotic alteration of the microbiome might provide an alternative to circumcision and a randomized clinical trial to compare the effect of antibiotics to circumcision on penile microbiome and HIV susceptibility in Ugandan men (NCT03412071) was just completed with analysis currently underway.

Comorbidities

Dr. Cara Wilson from the University of Colorado described her studies of host-microbe interactions and chronic immune activation. Dr. Wilson found that primary intestinal CD4+ T cells in vitro infected with HIV-1 and cultured in the presence of gut bacterial strain Prevotella stercorea displayed marked changes in the expression of genes involved in cellular proliferation and immune activation. 78 These changes included an upregulation of several genes related to the expression of granzymes, serine proteases that can induce apoptosis or promote an inflammatory response. 79,80 To investigate this association, the Wilson laboratory obtained samples from a previous clinical study^{81,82} and found that samples from people living with HIV had higher levels of activated CD4 T cells in the blood and colon and higher proportions of granzyme B-positive CD4 T cells in the gut. The levels of granzyme B-positive T cells appeared to correlate with the abundance of several bacterial taxa.

Interactions between diet and the gut microbiome appear to affect cardiovascular risk. ⁸³ For example, trimethylamine, a bacterial metabolite produced after meat, egg, and dairy consumption, is oxidized in the liver to trimethylamine N-oxide (TMAO) and appears to induce atherosclerosis. ⁸⁴ TMAO can be used as a biomarker of atherosclerosis and heart failure.

Dr. Marius Trøseid from the University of Oslo discussed his laboratories' work to study TMAO and metabolic syndromes in people living with HIV. In a cross-sectional study of people living with HIV, he saw little association between TMAO and myocardial infarction perhaps due to an overall increase of TMAO after initiation of protease inhibitor antiviral therapy.⁸⁹ Dr. Trøseid and collaborators have recently completed a Copenhagen-Oslo HIV Co-Morbidity and Microbiota Study (COMicS) to assess the links between HIV, sexual practice, and metabolic conditions. 90 They found contrasting effects of sexual practice and HIV infection on gut microbial diversity with MSM tending to have higher diversity, whereas people living with HIV tended to have less diversity resulting in MSM living with HIV having an intermediate diversity. They also identified several taxa previously linked to metabolic syndromes that were associated with HIV. 91 Regulation of gut microbiota by T cells has been proposed to reduce obesity⁹² and the COMicS study suggested a link between CD4 count and the amount of visceral adipose tissue and metabolic syndromes. 90 The laboratory

has also found soluble biomarkers of IL-1 activation to be predictive of myocardial infarction in people living with HIV, ⁹³ has investigated the association of acylation of gutderived lipopolysaccharides with inflammation, ⁹⁴ and is currently exploring the effects of microbial translocation on extracellular vesicles.

Vaccines and Therapeutics

Interactions between the microbiome and the host immune response^{95,96} and HIV and the microbiome^{91,97,98} suggest the potential for the microbiome to modulate HIV vaccine response. Dr. Roger Paredes from the IrsiCaixa AIDS Research Institute discussed his work to characterize the effects of the microbiome on vaccination response in clinical trials and mouse models. In data collected during the CUTHIVAC 03 (Cutaneous and Mucosal HIV Vaccination) trial, testing a modified vaccinia virus Ankara (MVA) HIV vaccine, Dr. Paredes and colleagues found that the expression of certain genes in whole blood and microbiome abundances in stool and skin swabs were potential predictors of the development of neutralizing antibodies.⁹⁹ In a BCN02-Romi trial attempting to reverse viral latency and eradicate infected cells ("kick and kill") using a combination of MVA vaccination and inducing agent romidepsin, 3 of 13 people living with HIV exhibited control of virus replication (viral load lower than 2000 copies/ml) for the entire duration of the 32-week treatment interruption and potential correlates of control were observed in the microbiome. 100 In a mouse model of multiple vaccinations, potential effects of sex and antibiotics on response to vaccination were observed. Further investigation into these areas is underway in the recently initiated MISTRAL (Microbiome-based stratification of individuals at risk of HIV-1 acquisition, chronic clinical complications, antimicrobial drug resistance, and unresponsiveness to therapeutic HIV-1 vaccination) project.

Dr. Alessandra Borgognone from the IrsiCaixa AIDS Research Institute continued the discussion of the BCN02-Romi trial of MVA vaccination and romidepsin in people living with HIV. She studied the stool bacterial abundances, peripheral blood mononuclear cell gene expression, and plasma protein composition of 3 individuals who controlled viral replication after treatment interruption and 10 individuals who did not control. Controllers appeared to have higher expression of genes related to immune activation and inflammatory response and an enrichment in protein markers of inflammation. Dr. Borgognone further found that controllers seemed to have higher levels of *Bacteroidales* and a depletion of *Clostridiales* bacteria, suggesting a potential role of the microbiome in vaccine response and subsequent control of HIV replication.

In addition to decreasing disease morbidity and mortality, vaccines can be used to probe the immune system. ¹⁰¹ Dr. Bali Pulendran from Stanford University discussed his research to better understand the human immune system utilizing vaccines. Vaccinations offer several benefits, including a defined time of immune perturbation allowing targeted high-throughput analysis, access to large and diverse populations receiving vaccinations, and an array of vaccine and adjuvant technologies providing varied immune stimuli. For example, in a study of the yellow fever vaccine (YF-17D), Dr. Pulendran and colleagues identified signatures of the vaccine-induced

innate immune response that predicted subsequent adaptive immune response. ¹⁰² Analysis of the metabolomic, transcriptomic, and cytokine profiles of the response to shingles vaccine Zostavax revealed additional insights. 103 Studies of vaccination also revealed interesting links to the microbiome. For example, the development of an adaptive response to influenza vaccination could be predicted by early molecular signatures, including increased expression of TLR5, a toll-like receptor, which senses flagellin from bacteria and triggers inflammation. 104 This unexpected link to bacteria was clarified by the finding that knockout mice lacking TLR5, along with antibiotic-treated or germ-free mice, exhibited reduced response to influenza vaccination. 105 A clinical trial testing the effects of antibiotic treatment before influenza vaccination (NCT02154061) revealed little effect in humans. However, a second trial focused on individuals with low pre-existing antibody titers showed a strong decrease in antibody titer in antibiotic-treated individuals ¹⁰⁶ revealing a critical role of microbiome in immune response.

Anxiety disorders and depression are considerably more prevalent in people living with HIV than among the general population ^{107,108} and a link between the gut microbiome and brain has been identified in anxiety disorders. ^{109,110} Dr. Shamsudheen Moidunny from the University of Miami and collaborators observed that both HIV and morphine could induce gut dysbiosis in a humanized mouse model ¹¹¹ and that anxiety-like behaviors increased with morphine treatment in a Tg26 mouse model expressing HIV proteins. Thus, the Tg26 mouse model may provide an important method to study interactions among HIV, microbiome, and opioid use. ¹¹²

Posters

Several investigators also presented posters summarizing research on HIV, the microbiome or related topics for online discussion of their research. Dr. Matthew Olaniyan from Edo University Iyamho looked at the co-occurrence of mosquito-borne parasites and HIV infection and links to markers of inflammation. Annette Aldous from George Washington University studied the effects of contraceptive use on the vaginal microbiome of adolescent girls. Dr. Alessandro Lazzaro from the University of Turin documented changes in the microbiome of people living with HIV in Zimbabwe before and after initiation of ART. Casey Martin from the University of Colorado Denver reported on the gut microbiomes of children infected at birth with HIV. Dickens Mahwayo from the Given-Secret Foundation discussed the need for an awareness campaign for vaginal ring pre-exposure prophylaxis in Malawi. Dr. Charlotte-Eve Short from the Imperial College London compared the microbiome composition in cervicovaginal secretions collected with menstrual cups with those collected by high vaginal swab.

Conclusion

The research presented at the sixth meeting of the International Workshop on Microbiome in HIV Pathogenesis, Prevention, and Treatment highlighted the broad interdisciplinary nature and promising progress being made in this area. Linkages between the microbiomes of multiple body sites and HIV transmission and pathogenesis were reported.

The multiple clinical trials presented show that microbiome research continues to move from observational studies into targeted alterations aimed to directly help patients. Continued collaborations between this broad coalitions of researchers and the communities of people living with HIV promise further improvements in understanding, diagnosing, and treating HIV-1 and its comorbidities.

References

- 1. Williams B, Mirmonsef P, Boucher CAB, *et al.*: A summary of the first HIV microbiome workshop 2015. AIDS Res Hum Retroviruses 2016;32:935–941.
- 2. Williams B, Ghosh M, Boucher CAB, *et al.*: A summary of the second annual HIV microbiome workshop. AIDS Res Hum Retroviruses 2017;33:1258–1264.
- 3. Williams B, Boucher C, Bushman F, *et al.*: A summary of the third annual HIV microbiome workshop. AIDS Res Hum Retroviruses 2018;34:828–834.
- Williams B, Ghosh M, Boucher CAB, et al.: A summary of the fourth annual virology education HIV microbiome workshop. AIDS Res Hum Retroviruses 2020;36:349– 356
- Sherrill-Mix S, Connors K, Aldrovandi GM, et al.: A summary of the fifth annual Virology Education HIV Microbiome workshop. AIDS Res Hum Retroviruses 2020;36:886–895.
- Van Belleghem J, Dabrowska K, Vaneechoutte M, Barr J, Bollyky P: Interactions between bacteriophage, bacteria, and the mammalian immune system. Viruses 2018; 11:10.
- 7. Zhang L, Forst CV, Gordon A, *et al.*: Characterization of antibiotic resistance and host-microbiome interactions in the human upper respiratory tract during influenza infection. Microbiome 2020;8:39.
- 8. Doudna JA, Charpentier E: The new frontier of genome engineering with CRISPR-Cas9. Science 2014;346: 1258096.
- Gulino K, Rahman J, Badri M, Morton J, Bonneau R, Ghedin E: Initial mapping of the New York City wastewater virome. mSystems 2020;5:e00876-19.
- Morrison CS, Chen P-L, Kwok C, et al.: Hormonal contraception and the risk of HIV acquisition: An individual participant data meta-analysis. PLoS Med 2015;12: e1001778.
- 11. Evidence for Contraceptive Options and HIV Outcomes (ECHO) Trial Consortium: HIV incidence among women using intramuscular depot medroxyprogesterone acetate, a copper intrauterine device, or a levonorgestrel implant for contraception: A randomised, multicentre, open-label trial. Lancet 2019;394:303–313.
- 12. Hapgood JP: ECHO: Context and limitations. Lancet 2020;395:e22.
- 13. Baeten JM, Kiarie J, Mastro TD, Mugo NR, Rees H: ECHO: Context and limitations—Authors' reply. Lancet 2020;395:e27.
- Miguel RDV, Calla NEQ, Aceves KM, Lopez FCD, Cherpes TL: ECHO: Context and limitations. Lancet 2020;395:e21.
- 15. Jewell BL, Smith JA, Padian NS, *et al.*: ECHO: Context and limitations. Lancet 2020;395:e25–e26.
- Hapgood JP, Kaushic C, Hel Z: Hormonal contraception and HIV-1 acquisition: Biological mechanisms. Endocr Rev 2018;39:36–78.

17. Thurman AR, Schwartz JL, Ravel J, *et al.*: Vaginal microbiota and mucosal pharmacokinetics of tenofovir in healthy women using tenofovir and tenofovir/levonorgestrel vaginal rings. PLoS One 2019;14:e0217229.

- Yang L, Hao Y, Hu J, et al.: Differential effects of depot medroxyprogesterone acetate administration on vaginal microbiome in hispanic white and black women. Emerg Microbes Infect 2019;8:197–210.
- Haddad LB, Wall KM, Kilembe W, et al.: Bacterial vaginosis modifies the association between hormonal contraception and HIV acquisition. AIDS 2018;32:595–604.
- Wessels JM, Felker AM, Dupont HA, Kaushic C: The relationship between sex hormones, the vaginal microbiome and immunity in HIV-1 susceptibility in women. Dis Model Mech 2018;11:dmm035147.
- 21. Abdool Karim Q, Abdool Karim SS, Frohlich JA, *et al.*: Effectiveness and safety of tenofovir gel, an antiretroviral microbicide, for the prevention of HIV infection in women. Science 2010;329:1168–1174.
- Noël-Romas L, Perner M, Molatlhegi R, et al.: Vaginal microbiome-hormonal contraceptive interactions associate with the mucosal proteome and HIV acquisition. PLoS Pathog 2020;16:e1009097.
- 23. Sullivan PS, Salazar L, Buchbinder S, Sanchez TH: Estimating the proportion of HIV transmissions from main sex partners among men who have sex with men in five US cities. AIDS 2009;23:1153–1162.
- Patel P, Borkowf CB, Brooks JT, Lasry A, Lansky A, Mermin J: Estimating per-act HIV transmission risk. AIDS 2014;28:1509–1519.
- 25. Kelley CF, Kraft CS, de Man TJB, *et al.*: The rectal mucosa and condomless receptive anal intercourse in HIV-negative MSM: Implications for HIV transmission and prevention. Mucosal Immunol 2017;10:996–1007.
- 26. Kelley CF, Pollack I, Yacoub R, et al.: Condomless receptive anal intercourse is associated with markers of mucosal injury in a cohort of men who have sex with men. SSRN Electron J 2020. DOI: 10.2139/ssrn.3710617.
- 27. Haaland RE, Fountain J, Hu Y, *et al.*: Repeated rectal application of a hyperosmolar lubricant is associated with microbiota shifts but does not affect PrEP drug concentrations: Results from a randomized trial in men who have sex with men. J Int AIDS Soc 2018;21:e25199.
- 28. Klase Z, Ortiz A, Deleage C, Mudd JC, *et al.*: Dysbiotic bacteria translocate in progressive SIV infection. Mucosal Immunol 2015;8:1009–1020.
- 29. Noguera-Julian M, Rocafort M, Guillén Y, *et al.*: Gut microbiota linked to sexual preference and HIV infection. EBioMedicine 2016;5:135–146.
- Armstrong AJS, Shaffer M, Nusbacher NM, et al.: An exploration of prevotella-rich microbiomes in HIV and men who have sex with men. Microbiome 2018; 6.10.1186/s40168-018-0580-7.
- Ortiz AM, Flynn JK, DiNapoli SR, et al.: Experimental microbial dysbiosis does not promote disease progression in SIV-infected macaques. Nat Med 2018;24:1313–1316.
- 32. Li SX, Sen S, Schneider JM, *et al.*: Gut microbiota from high-risk men who have sex with men drive immune activation in gnotobiotic mice and in vitro HIV infection. PLoS Pathog 2019;15:e1007611.
- Coleman SL, Neff PC, Li SX, et al.: Can gut microbiota of men who have sex with men influence HIV transmission? Gut Microbes 2020;11:610–619.

34. Lozupone CA, Li M, Campbell TB, *et al.*: Alterations in the gut microbiota associated with HIV-1 infection. Cell Host Microbe 2013;14:329–339.

- Ray S, Narayanan A, Giske CG, Neogi U, Söennerborg A, Nowak P: Altered gut microbiome under antiretroviral therapy: Impact of efavirenz and zidovudine. ACS Infect Dis 2021;7:1104–1115.
- 36. Pimentel M, Lembo A, Chey WD, *et al.*: Rifaximin therapy for patients with irritable bowel syndrome without constipation. N Engl J Med 2011;364:22–32.
- Arnold M, Sierra MS, Laversanne M, Soerjomataram I, Jemal A, Bray F: Global patterns and trends in colorectal cancer incidence and mortality. Gut 2016;66:683–691.
- Virostko J, Capasso A, Yankeelov TE, Goodgame B: Recent trends in the age at diagnosis of colorectal cancer in the US National Cancer Data Base, 2004–2015. Cancer 2019;125:3828–3835.
- 39. Wirbel J, Pyl PT, Kartal E, *et al.*: Meta-analysis of fecal metagenomes reveals global microbial signatures that are specific for colorectal cancer. Nat Med 2019;25:679–689.
- 40. Thomas AM, Manghi P, Asnicar F, *et al.*: Metagenomic analysis of colorectal cancer datasets identifies cross-cohort microbial diagnostic signatures and a link with choline degradation. Nat Med 2019;25:667–678.
- 41. Wu S, Rhee K-J, Albesiano E, *et al.*: A human colonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. Nat Med 2009;15: 1016–1022.
- 42. Housseau F, Wu S, Wick EC, *et al.*: Redundant innate and adaptive sources of IL17 production drive colon tumorigenesis. Cancer Res 2016;76:2115–2124.
- 43. Chung L, Thiele Orberg E, Geis AL, *et al.*: *Bacteroides fragilis* toxin coordinates a pro-carcinogenic inflammatory cascade via targeting of colonic epithelial cells. Cell Host Microbe 2018;23:203.e5–214.e5.
- Allen J, Hao S, Sears CL, Timp W: Epigenetic changes induced by *Bacteroides fragilis* toxin. Infect Immun 2019; 87:e00447-18.
- 45. Allen J, Sears CL: Impact of the gut microbiome on the genome and epigenome of colon epithelial cells: Contributions to colorectal cancer development. Genome Med 2019;11:11.
- Dejea CM, Wick EC, Hechenbleikner EM, et al.: Microbiota organization is a distinct feature of proximal colorectal cancers. Proc Natl Acad Sci U S A 2014;111: 18321–18326.
- 47. Johnson CH, Dejea CM, Edler D, *et al.*: Metabolism links bacterial biofilms and colon carcinogenesis. Cell Metab 2015;21:891–897.
- 48. Drewes JL, White JR, Dejea CM, *et al.*: High-resolution bacterial 16s rRNA gene profile meta-analysis and biofilm status reveal common colorectal cancer consortia. NPJ Biofilms Microbiomes 2017;3:34.
- 49. Tomkovich S, Dejea CM, Winglee K, *et al.*: Human colon mucosal biofilms from healthy or colon cancer hosts are carcinogenic. J Clin Invest 2019;129:1699–1712.
- Dejea CM, Fathi P, Craig JM, et al.: Patients with familial adenomatous polyposis harbor colonic biofilms containing tumorigenic bacteria. Science 2018;359:592–597.
- Zhang J, Haines C, Watson AJM, et al.: Oral antibiotic use and risk of colorectal cancer in the United Kingdom, 1989-2012: A matched case-control study. Gut 2019;68: 1971–1978.

- 52. Tuddenham SA, Koay WLA, Zhao N, White JR, Ghanem KG, Sears CL; H.I.V. Microbiome Re-analysis Consortium. The impact of human immunodeficiency virus infection on gut microbiota α-diversity: An individual-level meta-analysis. Clin Infect Dis 2020;70:615–627.
- Sinha R, Abu-Ali G, Vogtmann E, et al.: Assessment of variation in microbial community amplicon sequencing by the Microbiome Quality Control (MBQC) project consortium. Nat Biotechnol 2017;35:1077–1086.
- 54. Vermeulen R, Schymanski EL, Barabási AL, Miller GW: The exposome and health: Where chemistry meets biology. Science 2020;367:392–396.
- 55. Atashili J, Poole C, Ndumbe PM, Adimora AA, Smith JS: Bacterial vaginosis and HIV acquisition: A meta-analysis of published studies. AIDS 2008;22:1493–1501.
- Anahtar MN, Byrne EH, Doherty KE, et al.: Cervicovaginal bacteria are a major modulator of host inflammatory responses in the female genital tract. Immunity 2015;42:965–976.
- 57. Gosmann C, Anahtar MN, Handley SA, et al.: Lactobacillus-deficient cervicovaginal bacterial communities are associated with increased HIV acquisition in young South African women. Immunity 2017;46:29–37.
- 58. Bukusi E, Thomas KK, Nguti R, *et al.*: Topical penile microbicide use by men to prevent recurrent bacterial vaginosis in sex partners: A randomized clinical trial. Sex Transm Dis 2011;38:483–489.
- McClelland RS, Balkus JE, Lee J, et al.: Randomized trial of periodic presumptive treatment with high-dose intravaginal metronidazole and miconazole to prevent vaginal infections in HIV-negative women. J Infect Dis 2015; 211:1875–1882.
- 60. Francis SC, Looker C, Vandepitte J, *et al.*: Bacterial vaginosis among women at high risk for HIV in Uganda: High rate of recurrent diagnosis despite treatment. Sex Transm Infect 2016;92:142–148.
- 61. Hemmerling A, Harrison W, Schroeder A, *et al.*: Phase 1 dose-ranging safety trial of *Lactobacillus crispatus* CTV-05 for the prevention of bacterial vaginosis. Sex Transm Dis 2009;36:564–569.
- 62. Hemmerling A, Harrison W, Schroeder A, *et al.*: Phase 2a study assessing colonization efficiency, safety, and acceptability of *Lactobacillus crispatus* CTV-05 in women with bacterial vaginosis. Sex Transm Dis 2010;37:745–750
- Cohen CR, Wierzbicki MR, et al.: Randomized trial of Lactin-V to prevent recurrence of bacterial vaginosis. N Engl J Med 2020;382:1906–1915.
- 64. Lagenaur LA, Hemmerling A, Chiu C, *et al.*: Connecting the dots: Translating the vaginal microbiome into a drug. J Infect Dis 2021;223 (Supplement_3):S296–S306.
- 65. Hearps AC, Tyssen D, Srbinovski D, et al.: Vaginal lactic acid elicits an anti-inflammatory response from human cervicovaginal epithelial cells and inhibits production of pro-inflammatory mediators associated with HIV acquisition. Mucosal Immunol 2017;10:1480–1490.
- 66. Delgado-Diaz DJ, Tyssen D, Hayward JA, Gugasyan R, Hearps AC, Tachedjian G: Distinct immune responses elicited from cervicovaginal epithelial cells by lactic acid and short chain fatty acids associated with optimal and non-optimal vaginal microbiota. Front Cell Infect Microbiol 2020;9:446.
- 67. McMillan A, Rulisa S, Sumarah M, *et al.*: A multiplatform metabolomics approach identifies highly specific

- biomarkers of bacterial diversity in the vagina of pregnant and non-pregnant women. Sci Rep 2015;5:14174.
- 68. Srinivasan S, Morgan MT, Fiedler TL, et al.: Metabolic signatures of bacterial vaginosis. mBio 2015;6:e00204-15.
- Yeoman CJ, Thomas SM, Berg Miller ME, et al.: A multiomic systems-based approach reveals metabolic markers of bacterial vaginosis and insight into the disease. PLoS One 2013;8:e56111.
- Vitali B, Cruciani F, Picone G, Parolin C, Donders G, Laghi L: Vaginal microbiome and metabolome highlight specific signatures of bacterial vaginosis. Eur J Clin Microbiol Infect Dis 2015;34:2367–2376.
- Halperin DT, Bailey RC: Male circumcision and HIV infection: 10years and counting. Lancet 1999;354:1813–1815.
- 72. Auvert B, Taljaard D, Lagarde E, Sobngwi-Tambekou J, Sitta R, Puren A: Randomized, controlled intervention trial of male circumcision for reduction of HIV infection risk: The ANRS 1265 trial. PLoS Med 2005;2:e298.
- 73. Gray RH, Kigozi G, Serwadda D, Makumbi F, *et al.*: Male circumcision for HIV prevention in men in Rakai, Uganda: A randomised trial. Lancet 2007;369:657–666.
- Bailey RC, Moses S, Parker CB, et al.: Male circumcision for HIV prevention in young men in Kisumu, Kenya: A randomised controlled trial. Lancet 2007;369:643–656.
- 75. Prodger JL, Gray RH, Shannon B, *et al.*: Chemokine levels in the penile coronal sulcus correlate with HIV-1 acquisition and are reduced by male circumcision in Rakai, Uganda. PLoS Pathog 2016;12:e1006025.
- Liu CM, Hungate BA, Tobian AAR, et al.: Male circumcision significantly reduces prevalence and load of genital anaerobic bacteria. mBio 2013;4:e00076.
- 77. Liu CM, Prodger JL, Tobian AAR, *et al.*: Penile anaerobic dysbiosis as a risk factor for HIV infection. mBio 2017;8: e00996-17.
- 78. Yoder AC, Guo K, Dillon SM, *et al.*: The transcriptome of HIV-1 infected intestinal CD4+ T cells exposed to enteric bacteria. PLoS Pathog 2017;13:e1006226.
- 79. Hiebert PR, Granville DJ: Granzyme B in injury, inflammation, and repair. Trends Mol Med 2012;18:732–741
- 80. Wensink AC, Hack CE, Bovenschen N: Granzymes regulate proinflammatory cytokine responses. J Immunol 2015;194:491–497.
- 81. Dillon SM, Lee EJ, Kotter CV, *et al.*: Gut dendritic cell activation links an altered colonic microbiome to mucosal and systemic T-cell activation in untreated HIV-1 infection. Mucosal Immunol 2015;9:24–37.
- 82. Dillon SM, Kibbie J, Lee EJ, *et al.*: Low abundance of colonic butyrate-producing bacteria in HIV infection is associated with microbial translocation and immune activation. AIDS 2017;31:511–521.
- 83. Trøseid M, Andersen GO, Broch K, Hov JR: The gut microbiome in coronary artery disease and heart failure: Current knowledge and future directions. EBioMedicine 2020;52:102649.
- 84. Koeth RA, Wang Z, Levison BS, *et al.*: Intestinal microbiota metabolism of l-carnitine, a nutrient in red meat, promotes atherosclerosis. Nat Med 2013;19:576–585.
- 85. Wilson Tang WH, Wang Z, Levison BS, *et al.*: Intestinal microbial metabolism of phosphatidylcholine and cardio-vascular risk. N Engl J Med 2013;368:1575–1584.
- 86. Trøseid M, Ueland T, Hov JR, *et al.*: Microbiota-dependent metabolite trimethylamine-N-oxide is associated with

disease severity and survival of patients with chronic heart failure. J Intern Med 2015;277:717–726.

- 87. Li XS, Obeid S, Klingenberg R, et al.: Gut microbiotadependent trimethylamine N-oxide in acute coronary syndromes: A prognostic marker for incident cardiovascular events beyond traditional risk factors. Eur Heart J 2017;38:814–824.
- 88. Trøseid M: Gut microbiota and acute coronary syndromes: Ready for use in the emergency room? Eur Heart J 2017; 38:825–827.
- 89. Haissman JM, Knudsen A, Hoel H, *et al.*: Microbiota-dependent marker TMAO is elevated in silent ischemia but is not associated with first-time myocardial infarction in HIV infection. J Acquir Immune Defic Syndr 2016;71: 130–136.
- 90. Gelpi M, Vestad B, Hansen SH, *et al.*: Impact of human immunodeficiency virus-related gut microbiota alterations on metabolic comorbid conditions. Clin Infect Dis 2020; 71:e359–e367.
- 91. Vujkovic-Cvijin I, Somsouk M: HIV and the gut microbiota: Composition, consequences, and avenues for amelioration. Curr HIV/AIDS Rep 2019;16:204–213.
- 92. Petersen C, Bell R, Klag KA, *et al.*: T cell-mediated regulation of the microbiota protects against obesity. Science 2019;365.
- 93. Hoel H, Ueland T, Knudsen A, *et al.*: Soluble markers of interleukin 1 activation as predictors of first-time myocardial infarction in HIV-infected individuals. J Infect Dis 2020;221:506–509.
- Storm-Larsen C, Stiksrud B, Eriksen C, et al.: Microbial translocation revisited: Targeting the endotoxic potential of gut microbes in HIV-infected individuals. AIDS 2019; 33:645–653.
- Zheng D, Liwinski T, Elinav E: Interaction between microbiota and immunity in health and disease. Cell Res 2020;30:492–506.
- Skelly AN, Sato Y, Kearney S, Honda K: Mining the microbiota for microbial and metabolite-based immunotherapies. Nat Rev Immunol 2019;19:305–323.
- 97. Brenchley JM, Schacker TW, *et al.*: CD T cell depletion during all stages of HIV disease occurs predominantly in the gastrointestinal tract. J Exp Med 2004;200:749–759.
- 98. Guillén Y, Noguera-Julian M, Rivera J, *et al.*: Low nadir CD4+ T-cell counts predict gut dysbiosis in HIV-1 infection. Mucosal Immunol 2018;12:232–246.
- 99. Gonçalves E, Guillén Y, Lama JR, *et al.*: Host transcriptome and microbiota signatures prior to immunization profile vaccine humoral responsiveness. Front Immunol 2021;12.
- Borgognone A, Noguera-Julian M, Oriol B, et al. Gut microbiome signatures linked to HIV-1 reservoir size and viremia control. bioRxiv 2021.10.1101/2021.10.03.462590.

 Pulendran B: Systems vaccinology: Probing humanity's diverse immune systems with vaccines. Proc Natl Acad Sci U S A 2014;111:12300–12306.

- Querec TD, Akondy RS, Lee EK, et al.: Systems biology approach predicts immunogenicity of the yellow fever vaccine in humans. Nat Immunol 2008;10:116–125.
- Li S, Sullivan NL, Rouphael N, et al.: Metabolic phenotypes of response to vaccination in humans. Cell 2017b; 169:862.e17–877.e17.
- Nakaya HI, Wrammert J, Lee EK, et al.: Systems biology of vaccination for seasonal influenza in humans. Nat Immunol 2011;12:786–795.
- 105. Oh JZ, Ravindran R, Chassaing B, et al.: TLR5-mediated sensing of gut microbiota is necessary for antibody responses to seasonal influenza vaccination. Immunity 2014;41:478–492.
- 106. Hagan T, Cortese M, Rouphael N, *et al.*: Antibiotics-driven gut microbiome perturbation alters immunity to vaccines in humans. Cell 2019;178:1313.e13–1328.e13.
- Basu S, Chwastiak LA, Bruce RD: Clinical management of depression and anxiety in HIV-infected adults. AIDS 2005;19:2057–2067.
- 108. Beer L, Tie Y, Padilla M, Shouse RL: Generalized anxiety disorder symptoms among persons with diagnosed HIV in the United States. AIDS 2019;33:1781–1787.
- 109. Bravo JA, Forsythe P, Chew MV, et al.: Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. Proc Natl Acad Sci 2011;108:16050–16055.
- Dinan TG, Cryan JF: Brain-gut-microbiota axis and mental health. Psychosom Med 2017;79:920–926.
- 111. Meng J, Banerjee S, Zhang L, et al.: Opioids impair intestinal epithelial repair in HIV-infected humanized mice. Front Immunol 2020;10:2999.
- 112. Moidunny S, Benneyworth MA, Titus DJ, *et al.*: Glycogen synthase kinase-3 inhibition rescues sex-dependent contextual fear memory deficit in human immunodeficiency virus-1 transgenic mice. Br J Pharmacol 2020;177:5658–5676.

Address correspondence to:
Scott Sherrill-Mix
Department of Microbiology
Perelman School of Medicine
University of Pennsylvania
424 Johnson Pavilion, 3610 Hamilton Walk
Philadelphia, PA 19104
USA

E-mail: shescott@upenn.edu